



An Analysis of the Correlation between Physiochemical Characteristics of Soil and the Morphological Characteristics of *Grewia optiva* Drummond in the Northwestern Himalayan Region

Jyoti Dhiman^{a++*}, H. P. Sankhyan^{a#}, Neerja Rana^{b†},
Parul Sharma^{c‡} and Shikha Thakur^{a‡}

^a Department of Tree Improvement and Genetic Resources, College of Forestry, Dr. YS Parmar University of Horticulture and Forestry, Nauni-Solan (HP), 173230, India.

^b Department of Basic Sciences, College of Forestry, Dr. YS Parmar University of Horticulture and Forestry, Nauni-Solan (HP), 173230, India.

^c Department of Biotechnology, College of Horticulture, Dr. YS Parmar University of Horticulture and Forestry, Nauni-Solan (HP), 173230, India.

Authors' contributions

This work was carried out in collaboration among all authors. Author JD did the survey, data collection of the manuscript. Author HPS draw the road map for data collection and research of the manuscript. Authors NR, PS and ST member of advisory committee, they provided laboratory guidance for testing soil samples and analysis of collected data of the manuscript. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJPSS/2023/v35i183436

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/103330>

Original Research Article

Received: 18/05/2023

Accepted: 25/07/2023

Published: 05/08/2023

⁺⁺Ph. D. student;

[#]Professor and Head;

[†]Associate Professor;

[‡]Assistant Professor;

*Corresponding author: E-mail: jyotid175@gmail.com;

ABSTRACT

Continued research on the physiochemical properties of soil is critical for the long-term maintenance of cropping systems, (including trees), in order to harness their economic benefits. The current study was carried out in the Department of Tree Improvement and Genetic Resources, COF, Nauni, Solan (H.P.) during the period 2020-2023 to quantify the impact of soil nutrient variation on the morphological characteristics of *Grewia optiva* Drummond in different districts with variable climate and heterogeneous soils. The impacts on growth parameters (tree height, crown spread, leaf traits) of selected populations of *Grewia optiva* Drummond statistically analyzed using Karl Pearson correlation coefficient. Two composite soil samples representative of the different populations were drawn from the two depths *i.e.*, 0-15 cm (surface layer) and 15-30 cm (subsurface layer). These samples were collected underneath the selected populations of *Grewia optiva* Drummond. The collected soil samples were tested using standard soil methods and results were analysed using OPSTAT software. There was highly positive correlation observed between leaf area and soil N (0.509), leaf area and SOC (0.407), leaf area and soil P (0.728) and leaf area and soil K (0.577). Leaf length showed a highly significant correlation with SOC (0.401), soil N (0.509), soil P (0.710), and soil K (0.592). The tree height (0.385), tree diameter (0.602), crown spread (N-S) (0.629), crown spread (E-W) (0.334), branch nodal length (0.436) and leaf width (0.470) showed a significant positive correlation with soil P. Soil K showed a significant positive correlation with tree height (0.774), tree diameter (0.645), crown spread (N-S) (0.576), crown spread (E-W) (0.314), branch nodal length (0.737) and leaf width (0.592). Soil pH demonstrated highly significant correlation with Leaf width (0.449). The correlation developed between tree morphological and soil characteristics will help in quantify the impact of different soil characteristics on tree and leaf morphometric characteristics. It will further help in identification and selection of superior genotypes of *Grewia optiva* for further propagation to get improved genetic gain and for production of quality planting material.

Keywords: *Grewia*; yield; fertilization; soil quality; nitrogen; crop production.

1. INTRODUCTION

Grewia optiva, often known as Biul/Bihul or Bhimal, is a plant belonging to the Tiliaceae family. This species is favoured by mountainous farmers in Uttarakhand, Himachal Pradesh, Nepal, and elsewhere for qualities like as palatability, rapid growth, ease of propagation, and fodder production" [1]. "It supplies fodder during the lean season when there is no alternative to green fodder. It possesses more than 70 (%) potential DM digestibility and 56.7 percent effective degradability, making it a great energy source for ruminants" [2,3]. "*Grewia* is a genus with around 150 species worldwide, 42 of which are located on the Indian subcontinent" [4].

"Soil consists of definite chemical, physical, mineralogical and biological properties, which provide a medium for plant growth" [5]. "The knowledge of physiochemical properties viz; organic carbon, available Nitrogen (N), Phosphorus (P_2O_5), Potassium (K_2O), pH, electrical conductivity, soil texture and bulk density of soil is also important to determine the available nutrient status in soil and to develop

specific fertilizer recommendations" [6]. "The soil organic matter content, electrical conductivity and pH regulates not only macronutrients (N,P,K) but also micronutrients (Zn, Fe, B and Cu) for better uptake in plants" [7] "The response of trees to increasing atmospheric CO_2 concentrations is often mediated by the availability of nutrients in the soil" [8]. "Whether terrestrial ecosystems, forests, cropland trees are sources or sinks for CO_2 and their growth will ultimately depend on interactions of the C cycle with the cycles of nutrients, especially nitrogen (N) and phosphorus (P)" [9]. "An increased production of exoenzymes has been found in several studies with CO_2 enrichment, and this effect has depended on the availability of N in the soil" [10,11,12]. "Nitrogen (N) is one of the most important biological elements for plants, agricultural crops and forest trees, because it is a component of amino acids, proteins, genetic materials, pigments, and other key organic molecules" [13,14,15]. "N has an irreplaceable role in organ construction, material metabolism, fruit yield, and the quality formation of fruit trees" [16]. "Soil nitrogen (N) deficiencies can affect the photosynthetic N-use efficiency (PNUE), mesophyll conductance (g_m), and leaf N

allocation" [17]. "Potassium (K) is used for flowering purpose, it is also required for building of protein, photosynthesis, fruit quality and reduction of diseases" [18]. "Potassium is an activator of dozens of important enzymes, such as protein synthesis, sugar transport, N and C metabolism, and photosynthesis. It plays an important role in the formation of yield and quality improvement" [19,20]. "Potassium has strong mobility in plants and plays an important role in regulating cell osmotic pressure and balancing the cations and anions in the cytoplasm" [21]. "Phosphorus (P) is also an essential plant nutrient for various tree growth functions" [22]. "Plants take up Phosphorus in its inorganic form as phosphate" [23]. Phosphorus limitation decreases the efficiency of plant respiration [24] and night respiration may increase along the N/P ratio. "Therefore, considering the importance of the physiochemical properties of the soil mentioned above, the present study was carried out to perform a correlation and nutritional analysis of the soil under the populations of *Grewia optiva* Drummond in Himachal Pradesh and Uttarakhand" [25].

2. MATERIALS AND METHODS

2.1 Site Description and Data collection

The present study was carried out in four altitudinal zones viz; 400 to 800 m, 801-1200 m, 1201-1600 m and 1601-2000 m above mean sea level (a m s l), of Himachal Pradesh (HP) and Uttarakhand (UK). In each altitudinal zone, five populations (4 from HP and 1 from UK) were selected. The total populations under study were twenty. On each site/population, four trees of 20-30 cm diameter class were marked accordingly. The soil samples were collected during the month of November, when the species was in the seed ripening and fodder lopping stage. Two composite soil samples representative of the different population were drawn from the two depths i.e., 0-15cm (Surface layer) and 15-30cm (subsurface layer). These samples were collected underneath the selected twenty populations of *Grewia optiva* Drummond. Each sample was air dried, grounded with wooden pestle and mortar, sieved through 2 mm sieve and stored in plastic containers. The soil samples were then analysed for morphological (soil colour), physiochemical (bulk density, particle density, porosity, pH, EC), and available nutrient status using standard methods Table 1.

2.2 Experimental Design

Total Sites/ Populations	: 20
No. of composite samples of soil from each site/Population	: 2 (at two depth: 0-15cm & 15-30cm)
Total number of soil composite samples /treatments	: 40(20× 2)
Soil Depth	: 0-15 & 15-30cm
Design	: RBD(Factorial)

2.3 Statistical Analysis

Karl pearson correlation coefficient ($p < 0.05$) used to find correlation between physiochemical properties of soil and morphological characteristics of selected population of *Grewia optiva* which had been recorded simultaneously.

3. RESULTS AND DISCUSSION

Karl Pearson's correlation coefficients (at 5% level of significance) for tree and leaf morphometric characteristics (of different populations of *G. optiva*) and physiochemical characteristics of soil were worked out. It was evident from Table 2 and Fig. 1 that leaf area showed a significant positive correlation with soil N (0.509), SOC (0.407), soil P (0.728), and soil K (0.577). Leaf length showed a highly significant correlation with SOC (0.401), soil N (0.509), soil P (0.710), and soil K (0.592). The tree height (0.385), tree diameter (0.602), crown spread (N-S) (0.629), crown spread (E-W) (0.334), branch nodal length (0.436) and leaf width (0.470) showed a significant positive correlation with soil P. Soil K showed a significant positive correlation with tree height (0.774), tree diameter (0.645), crown spread (N-S) (0.576), crown spread (E-W) (0.314), branch nodal length (0.737) and leaf width (0.592). Soil pH showed highly significant correlation with Leaf width (0.449). Similar results recorded in Douglas-fir by Brix and Ebell [31], which revealed that "fertilization with Nitrogen increased basal area increment, stem height, and branch length, leaf area, leaf length-width, the no. of leaves per shoot increased markedly". Wang et al [32] reported "increased specific leaf area (SLA) and leaf area index (LAI) with N fertilization". A study on *Fagus sylvatica* by Meier and Leuschner [33] reported "positive effects on leaf area and LAI in forests with increased in nitrogen availability". In a similar study by Herbert and Fownes [34] "in native *Metrosideros polymorpha* forest showed

Table 1. Soil parameters analyzed with their methods of measurements

Sr.No.	Soil Parameters	Analysis Method Used
1	Soil colour	Munsell soil colour chart
2	pH	Digital pH meter [26].
3	Organic carbon (%)	Chromic acid titration method [27]
4	Available Nitrogen (kg ha ⁻¹)	Micro Kjeldhal Method [28]
5	Available Phosphorus (kg ha ⁻¹)	0.5 M sodium bicarbonate (NaHCO ₃) at 8.5 pH [29]
6	Available K (kg ha ⁻¹)	Flame photometric method (1N NH ₄ OAC extractable) [30]
7.	EC (dSm ⁻¹)	Digital EC meter [26]

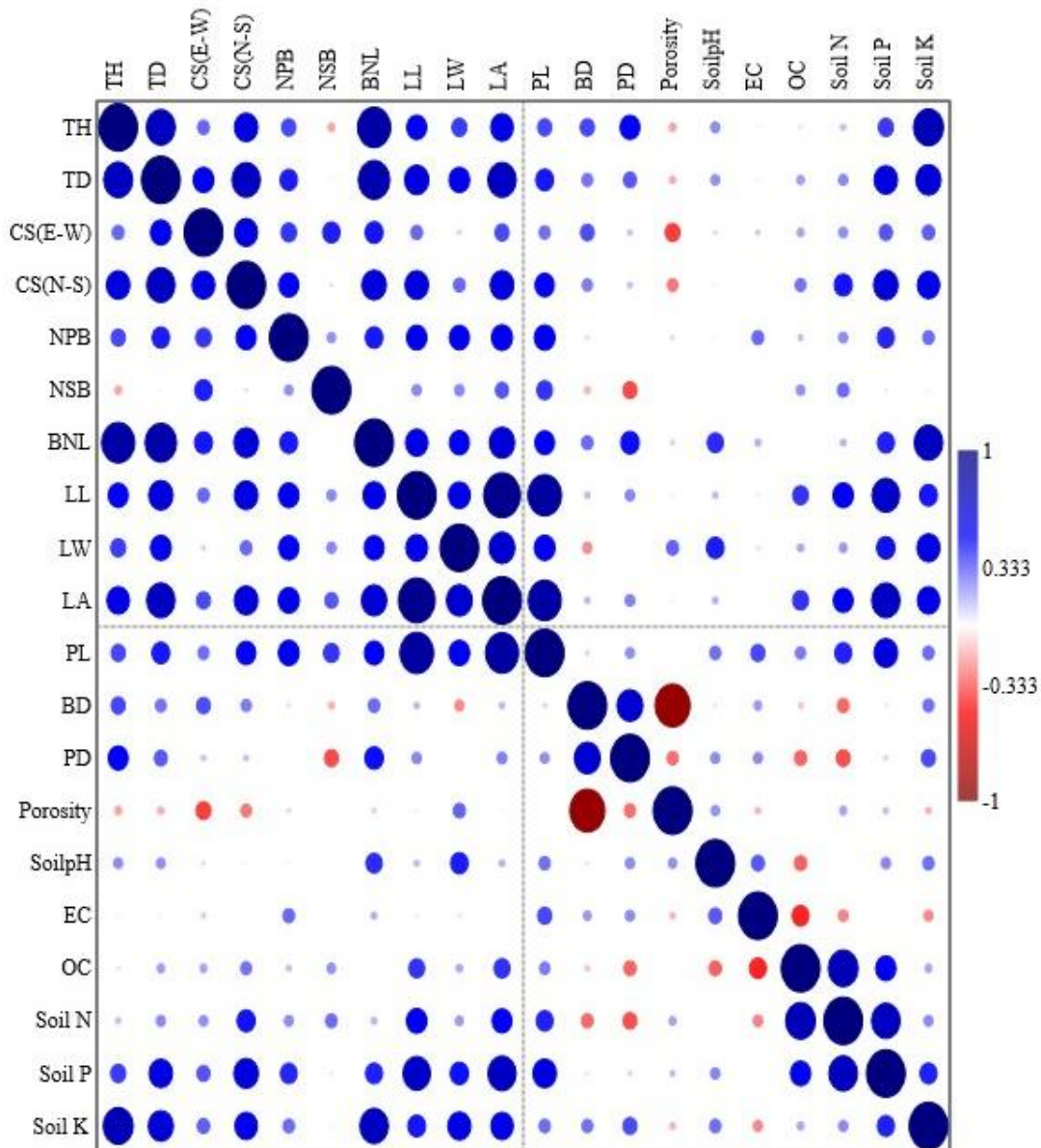


Fig. 1. correlation between physiochemical characteristics of soil and tree and leaf morphometric characteristics of *Grewia optiva drummond*

here TH-tree height, TD-tree diameter, CS-crown spread, NPB-number of primary branches, NSB-number of secondary branches, BNL-branch nodal length, LL-leaf length, LW-leaf width, LA-leaf area, PL-petiole length, BD-bulk density, PD-particle density, EC-electrical conductivity, OC-organic carbon, N-nitrogen, P-phosphorus, K-potassium

Table 2. Correlation between physiochemical characteristics of soil and tree and leaf morphometric characteristics of *Grewia optiva* drummond

Parameters	TH	TD	CS (E-W)	CS (N-S)	NPB	NSB	BNL	LL	LW	LA	PL	BD	PD	Porosity	Soil pH	EC	OC	Soil N	Soil P	Soil K	
TH	1																				
TD	0.747**	1																			
CS(E-W)	0.300	0.533**	1																		
CS(N-S)	0.602	0.728	0.588**	1																	
NPB	0.362	0.445	0.396	0.512**	1																
NSB	-0.178	0.026	0.442	0.065	0.215	1															
BNL	0.851	0.816	0.460	0.628	0.452	-0.016	1														
LL	0.512	0.623	0.298	0.605	0.522	0.230	0.572	1													
LW	0.383	0.531	0.090	0.294	0.517	0.236	0.513	0.562	1												
LA	0.581	0.732	0.358	0.610	0.541	0.332	0.655	0.944	0.665	1											
PL	0.362	0.456	0.281	0.495	0.534	0.400	0.503	0.867	0.534	0.852	1										
BD	0.361	0.273	0.348	0.257	-0.079	-0.155	0.291	0.136	-0.232	0.138	0.084	1									
PD	0.508	0.328	0.119	0.123	-0.023	-0.357	0.476	0.237	0.024	0.246	0.221	0.669	1								
Porosity	-0.179	-0.165	-0.380	-0.268	0.080	-0.003	-0.103	-0.043	0.307	-0.038	0.013	-0.903	-0.286	1							
SoilpH	0.225	0.221	0.078	0.037	0.028	0.008	0.416	0.135	0.449	0.148	0.282	-0.067	0.229	0.214	1						
EC	0.038	-0.042	0.105	-0.009	0.294	-0.011	0.154	0.046	-0.069	0.003	0.359	0.201	0.229	-0.145	0.328	1					
OC	0.062	0.192	0.178	0.273	0.127	0.217	-0.017	0.401**	0.169	0.407**	0.260	-0.120	-0.310	-0.018	-0.312	-0.433	1				
Soil N	0.129	0.230	0.220	0.464**	0.226	0.291	0.147	0.525**	0.202	0.509**	0.434**	-0.297	-0.350**	0.182	0.018	-0.244	0.770	1			
Soil P	0.385**	0.602**	0.334**	0.629**	0.429**	0.049	0.436**	0.710**	0.470**	0.728**	0.607**	-0.064	0.088	0.130	0.236	0.010	0.522	0.738	1		
Soil K	0.774**	0.645**	0.314**	0.576**	0.289	-0.043	0.737**	0.458**	0.592**	0.577**	0.287	0.277	0.356**	-0.149	0.284	-0.232	0.174	0.229	0.437	1	

**5% level of Significance, where TH-tree height, TD-tree diameter, CS-crown spread, NPB-number of primary branches, NSB-number of secondary branches, BNL-branch nodal length, LL-leaf length, LW-leaf width, LA-leaf area, PL-petiole length, BD-bulk density, PD-particle density, EC-electrical conductivity, OC-organic carbon, N-nitrogen, P-phosphorus, K-potassium

that increased available phosphorus promoted an increase in photosynthetic area which led to increased tree growth". A similar study in *Eucalyptus grandis* by Battie-Laclau et al [35] reported that "K and Na applications enhanced tree leaf area by increasing both leaf longevity and the mean area of individual leaves". "The most important role of pH is the control of nutrients solubility in soil. Nutrient availability usually decreases with increasing pH" [36]. "EC values affects uptake of nutrients by the tree. Very high or low EC decreases plants leaf size, leaf water content, leaf net photosynthetic rate (P_n), stomatal conductance (G_s), transpiration rate (T_r)", [37]. "Soil organic carbon is a natural resource for the sustainable development of human society and a key foundation for sustainable forestry development" [38]. "It plays an important role in the formation and conservation of soil structure, soil nutrient cycling and soil biodiversity. Nitrogen is considered to be the most important nutrient, and plants absorb more nitrogen than any other element. Nitrogen is essential in the formation of protein, and protein makes up much of the tissues of most living things. Increase in available phosphorus affected positively leaf area, crown spread, no. of branches and fruit. Phosphorus, is linked to a plant's ability to use and store energy, including the process of photosynthesis. It's also needed to help plants grow and develop normally" [39]. "Potassium is known to affect cell division, cell permeability formation of carbohydrates, translocation of sugars, various enzyme actions and resistance of some plants to certain diseases" [40,41]. It helps strengthen plants' abilities to resist disease and plays an important role in increasing crop yields and overall quality. Potassium also protects the plant when the weather is cold or dry, strengthening its root system and preventing wilt. Available soil Nitrogen (promotes leaf growth), Phosphorus (root, flower, and fruit), and Potassium supports stem and root growth and protein analysis.

4. CONCLUSION

The correlation developed between tree morphological and soil characteristics will help in quantifying the impact of different soil characteristics on tree and leaf morphometric characteristics and help in selection of superior populations, further improvement and fertilizers recommendation dose. This study will help in identification and selection of superior genotypes of *Grewia optiva* for further propagation to get

improved genetic gain and for production of quality planting material.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Mukherjee A, Modal T, Bist JK, Pattanayak A. Farmer's preference of fodder trees in mid hills of Uttarakhand: A comprehensive ranking using analytical hierarchy process. *Range Management and Agroforestry*. 2018;39:115-20.
2. Katoch R, Singh SK, Tripath A, Kumar N. Effect of seasonal variation in biochemical composition of leaves of fodder trees prevalent in the mid-hill region of Himachal Pradesh. *Range Management and Agroforestry*. 2017;38:234-40.
3. Singh B, Makkar PS, Negi SS. Rate and extent of digestion and potentially digestible dry matter and cell wall of various tree leaves. *Journal of Dairy Science*. 1989;72:3233-39.
4. Bhagta S, Thakur P, Sharma D. Genetic divergence study in *Grewia optiva* through Quantitative and molecular markers. *International Journal of Economic Plants*. 2021:029-033. Available:<https://doi.org/10.23910/2/2020.0396>
5. Thakre YG, Choudhary MD, Raut RD. Physicochemical characterization of red and black soils of Wardha region. *International Journal of Chemical and Physical Sciences*. 2012;1:60-66.
6. Sumithra S, Ankalaiah C, Rao D, Yamuna RT. A case study on physico-chemical characteristics of soil around industrial and agricultural area of yerraguntla, kadapa district, AP, India. *International Journal of Geology Earth and Environmental Sciences*. 2013;3:28-34.
7. Havlin JL. Soil: Fertility and nutrient management. In *landscape and land capacity*; Wang Y, Ed.; CRC Press Inc.: Boca Raton, FL, USA; 2020.
8. Schleppe P, Korner C, Klein T. Increased nitrogen availability in the soil under mature *Picea abies* trees exposed to elevated CO_2 concentrations. *Frontiers in Forests and Global Change*. 2019;2:59.
9. Ellsworth DS, Anderson IC, Crous KY, Cooke J, Drake JE, Gherlenda AN.

- Elevated CO₂ does not increase eucalypt forest productivity on a low-phosphorus soil. *Nature Climate Change*. 2017;7:279–283.
DOI: 10.1038/nclimate3235
10. Drake JE, Gallet-Budynek A, Hofmockel KS, Bernhardt ES, Billings SA, Jackson RB. Increases in the flux of carbon belowground stimulate nitrogen uptake and sustain the long-term enhancement of forest productivity under elevated CO₂. *Ecology Letters*. 2013;14:349–357.
DOI: 10.1111/j.1461-0248.2011.01593.x
 11. Meier IC, Finzi AC, Phillips RP. Root exudates increase N availability by stimulating microbial turnover of fast-cycling N pools. *Soil Biology and Biochemistry*. 2017;106:119–128.
DOI: 10.1016/j.soilbio.2016.12.004
 12. Ochoa-Hueso R, Hughes J, Delgado-Baquerizo M, Drake JE, Tjoelker MG, Pineiro J. Rhizosphere-driven increase in nitrogen and phosphorus availability under elevated atmospheric CO₂ in a mature *Eucalyptus* woodland. *Plant and Soil*. 2017;416:283–295.
DOI: 10.1007/s11104-017-3212-2
 13. Chen LH, Dong TF, Duan BL. Sex-specific carbon and nitrogen partitioning under N deposition in *Populus cathayana*. *Trees*. 2014;28:793–806.
Available: <https://doi.org/10.1007/s00468-014-0992-3>
 14. Ji DH, Mao QZ, Watanabe Y, Kitao M, Kitaoka S. Effect of nitrogen loading on the growth and photosynthetic responses of Japanese larch seedlings grown under different light regimes. *Journal of Agricultural Meteorology*. 2015;71:232–238
Available: <https://doi.org/10.2480/agrmet.D-14-00027>
 15. Liu N, Wang J, Guo Q, Wu S, Rao X, Cai X, Lin Z. Alterations in leaf nitrogen metabolism indicated the structural changes of subtropical forest by canopy addition of nitrogen. *Ecotoxicology and Environmental Safety*. 2018;160:134–143.
Available: <https://doi.org/10.1016/j.ecoenv.2018.05.037>
 16. Bai L, Deng H, Zhang X, Yu X, Li Y, Gibberellin Is Involved in Inhibition of cucumber growth and nitrogen uptake at suboptimal root-zone temperatures. *PLoS One*. 2016;11(5):e0156188.
Available: <https://doi.org/10.1371/journal.pone.0156188>
 17. Tang J, Sun B, Cheng R, Shi Z, Luo D, Liu S, Centritto. Effects of soil nitrogen (N) deficiency on photosynthetic N-use efficiency in N-fixing and non-N-fixing tree seedlings in subtropical China. *Scientific Reports*. 2019;9:4604.
Available: <https://doi.org/10.1038/s41598-019-41035-1>
 18. Valente DSM, Queiroz DM, Pinto F, Santos NT, Santos FL. Definition of Management zones in coffee production fields based on apparent soil electrical conductivity. *Scientia Agricola*. 2012;69: 173-179.
 19. Marschner H. *Marschner's Mineral Nutrition of Higher Plants*. Cambridge, MA: Academic press; 2012.
 20. Oosterhuis D, Loka D, Kawakami E, Pettigrew W. The physiology of potassium in crop production. *Advances in Agronomy*. 2014;126:203–234.
DOI: 10.1016/B978-0-12-800132-5.00003-1
 21. Hu W, Zhao W, Yang J, Oosterhuis DM, Loka DA, Zhou Z. Relationship between potassium fertilization and nitrogen metabolism in the leaf subtending the cotton (*Gossypium hirsutum* L.) boll during the boll development stage. *Plant Physiology and Biochemistry*. 2016a;101: 113–123.
DOI: 10.1016/j.plaphy.2016.01.019
 22. Jonard M, Furst A, Verstraeten A. Tree mineral nutrition is deteriorating in Europe. *Global Change Biology*. 2015;21:418-430.
 23. Becquer A, Trap J, Irshad U, Ali MA, Claude P. From soil to plant, the journey of P through trophic relationships and ectomycorrhizal association. *Frontiers in Plant Science*. 2014;5:548.
DOI: 10.3389/fpls.2014.00548
 24. Jiang M, Caldararu S, Zaehle S, Ellsworth DS, Medlyn BE. Towards a more physiological representation of vegetation phosphorus processes in land surface models. *New Phytologist* 2019;222:1223–1229.
DOI: 10.1111/nph.15688
 25. Sankhyan HP, Dhiman J, Chand K, Negi B. Soil nutrient analysis underneath *Grewia optiva* population. *International Journal of Bio-resource and Stress Management*. 2021;12(6):645-54.
 26. Jackson ML. *Soil Chemical Analysis*. Prentice Hall of India (Pvt.) Ltd., New Delhi. 1973;85:251-252.

27. Walkley AJ, Black IA. Estimation of soil organic carbon by the chromic acid titration method. *Soil Science*. 1934;37:29-38.
28. Subbaiah BV, Asija GL. A rapid method for the estimation of available nitrogen in soil. *Current Science*. 1956;25:258-260.
29. Olsen SR, Cole CV, Watanable FS, Dean LA. Estimation of available phosphorus in soil by extraction with NaHCO_3 , USDA Circular, US Washington. 1954;939:19.
30. Merwin HD, Peech M. Exchangeability of soil potassium in the sand, silt and clay fractions as influenced by the nature of the complementary exchangeable cation. *Soil Science Society of America Journal*. 1951;15:125-28.
31. Brix H, Ebell LF. Effects of Nitrogen fertilization on Growth, leaf area, and photosynthesis rate in Douglas-Fir. *Forest Sciences*. 1969;15:189-196.
32. Wang D, Mathew WM, Jindong S, Xiaohui F, Fernando M, Dokyoung L, Michael C. Impact of nitrogen allocation on growth and photosynthesis of *Miscanthus (Miscanthus X giganteus)* GCB *Bioenergy*. 2012;4: 688–697.
33. Meier IC, Leuschner C. Leaf size and leaf area index in *Fagus sylvatica* forests: Competing effects of precipitation, temperature, and nitrogen availability. *Ecosystem*. 2008;11:655–669.
34. Herbert DA, Fownes JH. Phosphorus limitation of forest leaf area and net primary production on a highly weathered soil. *Biogeochemistry*. 1995;29:223-235.
35. Battie-Laclau P, Laclau JP, Piccolo MC, Arenque Beri C, Mietton L., Muniz MR, Meille- Buckeridge MS, Nouvellon Y, Ranger J, Bouillet JP. Influence of potassium and sodium nutrition on leaf area components in *Eucalyptus grandis* trees. *Plant and Soil*. 2013;371:19–35.
36. Kazem H, Tayebbeh M, Farani S, Jamaati-e-Somarin. Effect of elemental sulphur and compost on pH, electrical conductivity and phosphorus availability of one clay soil. *African Journal of Biotechnology*. 2012;11: 1425-1432.
37. Sonneveld C, De Kreij C. Response of cucumber (*Cucumis sativus* L.) to an unequal distribution of salts in the root environment. *Plant and Soil*. 1996;209:45-56.
38. Pan G, Lu H, Li L, Zhang J, Zhang X. Soil carbon sequestration with bioactivity: A new emerging frontier for sustainable soil management. *Advances in Earth Science*. 2015;30:940–951.
39. Yosuf M, Li J, Lu J, Ren T, Cong R, Fahad S, Li X. Effects of fertilization on crop production and nutrient-supplying capacity under rice-oilseed rape rotation system. *Scientific Reports*. 2017;7:1-9.
40. Miller C, Turk LM. *Fundamentals of soil science biotech*. Books, 1123/74, Trinagar, Delhi, India. 2002:157.
41. Smith JL, Doran JW. Measurement and use of pH and electrical conductivity for soil quality analysis. *Methods for Assessing Soil Quality*. 1996:169-185.

© 2023 Dhiman et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/103330>